

Note

Synthesis of 6'-GM2, a regioisomer of ganglioside GM2, for studying the mechanism of action of GM2 activator¹

Hideharu Ishida^a, Yuko Ito^a, Eiji Tanahashi^a, Yu-Teh Li^b,
Makoto Kiso^{a,*}, Akira Hasegawa^{a,2}

^a Department of Applied Bioorganic Chemistry, Gifu University, Gifu 501-11, Japan

^b Department of Biochemistry, Tulane University School of Medicine New Orleans, LA 70112, USA

Received 6 March 1997; accepted 31 March 1997

Keywords: Ganglioside; GM2 activator; Tay–Sachs disease

GM2 activator is a protein cofactor that assists β -hexosaminidase (HexA) in hydrolyzing the β -linked *N*-acetylgalactosamine (GalNAc) in ganglioside GM2. Tay–Sachs disease can be caused by a deficiency of HexA or GM2 activator [2,3]. Although the GM2 activator was discovered in 1973 [4], the mechanism of action of this activation protein is still not well understood. In this paper, we report a synthesis of the regioisomer of GM2, 6'-GM2, in which the GalNAc residue is linked, not to the 4-OH, but to the 6-OH of Gal to show that the resistance of GalNAc in GM2 to HexA is due to the specific structure of the GM2-epitope, β -D-GalNAc-(1 \rightarrow 4)- α -Neu5Ac-(2 \rightarrow 3)- β -D-Gal \rightarrow .

Removal of the benzylidene group from 2-(trimethylsilyl)ethyl 2-*O*-benzyl-4,6-*O*-benzylidene-3-*O*-(4-methoxybenzyl)- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -D-glucopyranoside (**1**) [5] with aqueous 80% acetic acid at 50 °C gave, after column chromatography, the lactose acceptor **2** in 93% yield. The glycosylation of **2** with the oxazoline **3**, which

was prepared [6] from the peracetylated derivative of GalNAc, in the presence of pyridinium *p*-toluenesulfonate in dichloromethane for 24 h under reflux gave the desired β -glycoside **4** in 61% yield. Removal of the 4-methoxybenzyl group in **4** in acetonitrile–water in the presence of ceric ammonium nitrate (CAN) for 1 h at room temperature afforded the expected glycosyl acceptor **5** in 67% yield. Glycosylation [7] of **5** with the phenyl 2-thioglycoside of Neu5Ac (**6**) [8] in acetonitrile for 3 h at –25 °C in the presence of *N*-iodosuccinimide (NIS)-trifluoromethanesulfonic acid (TfOH) [9] gave the expected α -glycoside **7** in a low yield (23%), with the unreacted glycosyl acceptor being recovered in 70% yield. The observed chemical shift (δ 4.87) for H-4 in the Neu5Ac moiety is characteristic for the α -glycosidic linkage [10]. Removal of the benzyl groups from **7** by catalytic hydrogenolysis over 10% Pd–C in 1:1 ethanol–acetic acid for 24 h at 40 °C and subsequent acetylation gave the peracetylated tetrasaccharide **8** in 88% yield. The resulting tetrasaccharide was converted into the corresponding trichloroacetimidate **10** in good yield by selective removal of the 2-(trimethylsilyl)ethyl group with trifluoroacetic acid and subsequent imidate formation [11,12].

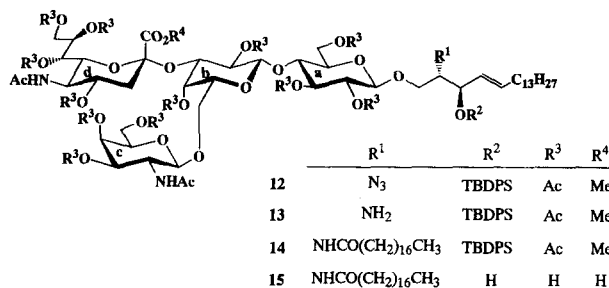
The final glycosylation [13] of (2*S*,3*R*,4*E*)-2-

* Corresponding author.

¹ Synthetic studies on sialoglyconjugates, Part 94. For Part 93, see ref. [1].

² Deceased 10 October 1996.

azido-3-*O*-(*tert*-butyldiphenylsilyl)-4-octadecene-1,3-diol (**11**) [14] with **10** in dichloromethane for 12 h at 0 °C in the presence of trimethylsilyl trifluoromethanesulfonate and 4 Å molecular sieves (AW-300) gave only the β -glycoside **12** in 80% yield. Selective reduction [15] of the azido group in **12** with triphenylphosphine in benzene–water gave the amine **13**, which on condensation [16] with octadecanoic acid using 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (WSC) in dichloromethane furnished the fully protected target compound **15** as follows: desilylation [17] of **14** by treatment with tetrabutylammonium fluoride in acetonitrile, and *O*-deacylation with sodium methoxide in methanol, and subsequent saponification of the methyl ester group, yielded the desired 6'-GM2 ganglioside **15** in a quantitative yield after a Sephadex LH-20 column chromatography.



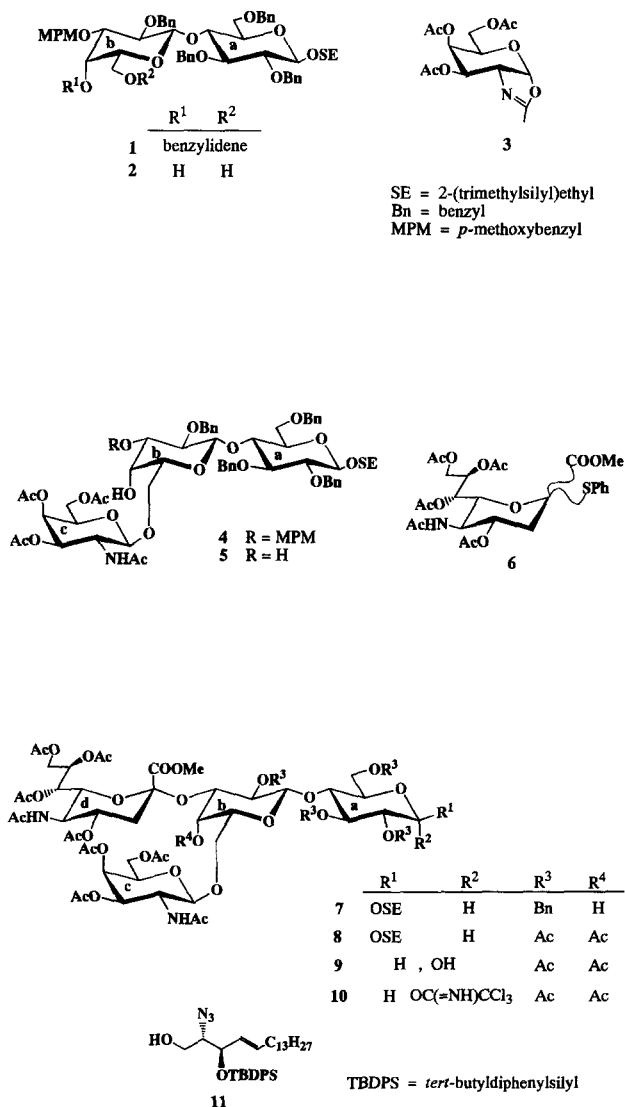
In contrast to GM2, the β -D-GalNAc-(1 → 6)-Gal linkage in 6'-GM2 was readily hydrolyzed by HexA in the absence of the GM2 activator, indicating that the resistance of GM2 to HexA is due to the specific structure of the GM2-epitope and that the GM2 activator may interact with GM2 to make GalNAc accessible to HexA.³

1. Experimental

General procedures.—Specific rotations were determined with a Union PM-201 polarimeter at 25 °C, and ¹H NMR spectra were recorded at 270 MHz with a Jeol JNM-GX 270 spectrometer or at 200 MHz with a Varian Gemini-2000 spectrometer. Preparative TLC was performed on Silica Gel 60 (E. Merck), and column chromatography on silica gel (Fuji Silysia Co., 300 mesh) was accomplished with the solvent systems (v/v) specified. Concentrations and evaporations were conducted in vacuo.

2-(Trimethylsilyl)ethyl 2-O-benzyl-3-O-(4-methoxybenzyl)- β -D-galactopyranosyl-(1 → 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (2).—A soln of **1** (1.01 g, 1 mmol) in aq 80% CH₃COOH (5 mL) was heated for 48 h at 40 °C and concd. Column chromatography (1:1 AcOEt–hexane) of the residue on silica gel gave **2** (0.74 g, 93%) as a syrup: [α]_D +21.7° (c 1.0, CH₂Cl₂); ¹H NMR (CDCl₃): δ 1.00 (m, 2 H, Me₃SiCH₂CH₂), 3.78 (s, 3 H, MeOPh), and 6.82–7.53 (m, 29 H, 6 Ph). Anal. Calcd for C₅₃H₆₆O₁₂Si (923.19): C, 68.96; H, 7.21. Found: C, 68.8; H, 7.11.

2-(Trimethylsilyl)ethyl 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl-(2 → 6)-2-O-benzyl-3-O-(4-methoxybenzyl)- β -D-galactopyranosyl-(1 → 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (4).—To a soln of 2-acetamido-1,3,4,6-tetra-O-acetyl-2-deoxy-D-glucopyranose (4.7 g, 12.0 mmol)



³ Details of the studies on the enzymatic hydrolysis of 6'-GM2 will be published elsewhere.

was added FeCl_3 (4.7 g, 29.0 mmol), and the mixture was stirred for 1.5 h at 50 °C, at the end of which time it was cooled and neutralized with M NaHCO_3 . The insoluble materials were removed by filtration and washed thoroughly with CH_2Cl_2 . The filtrate and washings were combined and extracted with CH_2Cl_2 . The extract was washed with M NaHCO_3 and brine, dried (Na_2SO_4), and concd to give the oxazoline **3** (3.85 g, 95%), which was used for the next reaction without further purification.

To a soln of **2** (3.6 g, 3.9 mmol) and **3** (3.85 g, 11.7 mmol) in CH_2Cl_2 (30 mL) was added pyridinium *p*-toluenesulfonate (0.59 g, 0.13 mmol), and the soln was stirred under reflux for 24 h. Dichloromethane (50 mL) was added, and the soln was successively washed with M NaHCO_3 , water, and 2 M HCl, dried (Na_2SO_4), and concd. Column chromatography (1:1 AcOEt–hexane) of the residue on silica gel gave **5** (3.0 g, 61%) as an amorphous mass: $[\alpha]_D +3.6^\circ$ (*c* 0.9, CH_2Cl_2); ^1H NMR (CDCl_3): δ 1.00 (m, 2 H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.96, 2.02, 2.04, 2.09 (4 s, 12 H, 3 AcO and AcN), 2.66 (t, 1 H, $J_{1,2} = J_{2,3} = 7.6$ Hz, H-2a), 3.81 (s, 3 H, MeOPh), 5.02 (d, 1 H, $J_{\text{NH},2} = 8.8$ Hz, NH), 6.83–7.55 (m, 24 H, 5 Ph). Anal. Calcd for $\text{C}_{67}\text{H}_{85}\text{NO}_{20}\text{Si}$ (1252.5): C, 64.25; H, 6.84; N, 1.12. Found: C, 64.20; H, 6.75; N, 0.83.

2-(Trimethylsilyl)ethyl 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl-(2 \rightarrow 6)-2-O-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (5).—A soln of **4** (0.66 g, 0.53 mmol) and ceric ammonium nitrate (CAN; 1.45 g, 2.6 mmol) in MeOH (80 mL) was stirred for 1 h at room temperature. After completion of the reaction, NaHCO_3 was added, and the mixture was concd and extracted with CH_2Cl_2 . The extract was washed with water, dried (Na_2SO_4), and concd. Column chromatography (45:1 CH_2Cl_2 –MeOH) of the residue on silica gel gave **6** (0.4 g, 67%) as an amorphous mass: $[\alpha]_D +4.5^\circ$ (*c* 0.44, CH_2Cl_2); ^1H NMR (CDCl_3): δ 1.00 (m, 2 H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.96, 1.97, 2.04, 2.10 (4 s, 12 H, 3 AcO and AcN), 2.92 (t, 1 H, $J_{1,2} = J_{2,3} = 5.8$ Hz, H-2a), 5.05 (d, 1 H, $J_{1,2} = 10.8$ Hz, H-1c), 5.08 (s, 1 H, H-4c), 5.12 (d, 1 H, $J_{2,3} = 8.8$ Hz, H-3c), 5.52 (d, 1 H, $J_{\text{NH},2} = 8.8$ Hz, NH), 7.26–7.53 (m, 20 H, 4 Ph). Anal. Calcd for $\text{C}_{59}\text{H}_{77}\text{NO}_{19}\text{Si}$ (1132.3): C, 62.58; H, 6.85; N, 1.24. Found: C, 62.56; H, 6.65; N, 1.21.

2-(Trimethylsilyl)ethyl 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl-(2 \rightarrow 6)-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow

3)]-2-O-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-glucopyranoside (7).—To a soln of **5** (100 mg, 0.09 mmol) and methyl (phenyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-thio-2-nonulopyranosid)onate (**6**; 103 mg, 0.1 mmol) in dry acetonitrile (1.7 mL) was added 3 Å molecular sieves (100 mg), and the mixture was stirred for 5 h at room temperature, then cooled to -25°C . To this mixture were added, with stirring, *N*-iodosuccinimide (NIS; 80 mg, 0.36 mmol) and trifluoromethanesulfonic acid (TfOH; 3.1 μL , 0.036 mmol), and the stirring was continued for 3 h at -25°C . The precipitate was removed by filtration and washed thoroughly with CH_2Cl_2 . The filtrate and washings were combined, and the soln was successively washed with M NaHCO_3 , M $\text{Na}_2\text{S}_2\text{O}_3$, and water, dried (Na_2SO_4), and concd. Column chromatography (40:1 CH_2Cl_2 –MeOH) of the residue on silica gel gave **7** (32 mg, 23%) as an amorphous mass: $[\alpha]_D -8.1^\circ$ (*c* 0.6, CH_2Cl_2); ^1H NMR (CDCl_3): δ 1.00 (m, 2 H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.88, 1.96, 1.97, 2.00, 2.01, 2.03, 2.04, 2.09, 2.11 (9 s, 27 H, 7 AcO and 2 AcN), 2.42 (dd, 1 H, $J_{\text{gem}} = 12.6$, $J_{3\text{eq},4} = 5.7$ Hz, H-3d_{eq}), 3.80 (s, 3 H, MeO), 4.66 (dd, 1 H, $J_{5,6} = 3.8$, $J_{\text{gem}} = 12.6$ Hz, H-6a), 4.87 (m, 1 H, H-4d), 5.02 (d, 1 H, $J_{1,2} = 9.5$ Hz, H-1c), 5.04 (s, 1 H, H-3c), 5.12 (d, 1 H, $J_{2,3} = 12.1$ Hz, H-3c), 5.34 (dd, 1 H, $J_{6,7} = 2.2$, $J_{7,8} = 9.5$ Hz, H-7d), 5.35 (m, 1 H, H-8d), 5.63 (d, 1 H, $J_{\text{NH},2} = 9.5$ Hz, NH), 7.24–7.55 (m, 20 H, 4 Ph). Anal. Calcd for $\text{C}_{79}\text{H}_{104}\text{N}_2\text{O}_{31}\text{Si}$ (1605.8): C, 59.09; H, 6.53; N, 1.74. Found: C, 58.93; H, 6.31; N, 1.45.

2-(Trimethylsilyl)ethyl 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl-(2 \rightarrow 6)-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-2,4-di-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-acetyl- β -D-glucopyranoside (8).—A soln of **7** (0.89 g, 0.55 mmol) in EtOH (23 mL) and CH_3COOH (23 mL) was hydrogenated in the presence of 10% Pd–C (0.9 g) for 24 h at 40 °C, then filtered and concd. The residue was acetylated with a mixture of Ac_2O (6 mL) and pyridine (9.8 mL) for 12 h at 40 °C. The product was purified by chromatography on a column of silica gel with 20:1 CH_2Cl_2 –MeOH to give **8** (0.71 g, 88%) as an amorphous mass: $[\alpha]_D -19.0^\circ$ (*c* 2.0, CH_2Cl_2); ^1H NMR (CDCl_3): δ 1.00 (m, 2 H, $\text{Me}_3\text{SiCH}_2\text{CH}_2$), 1.69 (t, 1 H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.3$ Hz, H-3d_{ax}), 1.85–2.19 (14 s, 42 H, 12 AcO and 2 AcN), 2.56 (dd, 1 H, $J_{3\text{eq},4} = 4.6$ Hz, H-3d_{eq}), 3.95 (d, 1 H, $J_{1,2} = 9.7$ Hz, H-1b), 4.50 (d, 1 H, $J_{1,2} = 8.1$ Hz, H-1a), 4.82 (d, 1 H, $J_{1,2} = 6.6$ Hz, H-1c), 4.97

(dd, 1 H, $J_{1,2} = J_{2,3} = 10.2$ Hz, H-2a), 5.12 (t, 1 H, $J_{2,3} = J_{3,4} = 8.8$ Hz, H-3a), 5.19 (d, 1 H, $J_{\text{NH},2} = 8.6$ Hz, NH), 5.33 (dd, 1 H, $J_{6,7} = 2.7$, $J_{7,8} = 8.6$ Hz, H-7d), 5.41 (d, 1 H, $J_{3,4} = 3.3$ Hz, H-4b), 5.64 (m, 1 H, H-8d), 5.76 (dd, 1 H, $J_{2,3} = 11.4$, $J_{3,4} = 3.3$ Hz, H-3c), 6.71 (d, 1 H, $J_{\text{NH},2} = 7.1$ Hz, NH). Anal. Calcd for $\text{C}_{61}\text{H}_{90}\text{N}_2\text{O}_{36}\text{Si}$ (1455.4): C, 50.34; H, 6.23; N, 1.92. Found: C, 50.23; H, 6.19; N, 1.80.

2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl-(2 \rightarrow 6)-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-2,4-di-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-acetyl-D-glucopyranose (9).—To a soln of **8** (0.56 g, 0.38 mmol) in dry CH_2Cl_2 (2.7 mL) was added CH_3COOH (2.6 mL), and the soln was stirred for 2 h at room temperature and then concd. Column chromatography (15:1 CH_2Cl_2 –MeOH) of the residue on silica gel gave **9** (0.52 g, quantitative) as an amorphous mass: ^1H NMR (CDCl_3): δ 1.70 (t, 1 H, $J_{\text{gem}} = J_{3\text{ax},4} = 10.8$ Hz, H-3dax), 1.86–2.28 (14 s, 42 H, 12 AcO and 2 AcN), 2.57 (dd, 1 H, $J_{3\text{eq},4} = 4.7$ Hz, H-3deq). Anal. Calcd for $\text{C}_{61}\text{H}_{90}\text{N}_2\text{O}_{36}$ (1355.2): C, 49.63; H, 5.8; N, 2.07. Found: C, 49.54; H, 5.63; N, 1.84.

2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl-(2 \rightarrow 6)-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-2,4-di-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-acetyl- α -D-glucopyranosyl trichloroacetimidate (10).—To a soln of **9** (0.47 g, 0.35 mmol) in CH_2Cl_2 (8 mL) and trichloroacetonitrile (1.45 mL) cooled to 0 °C was added 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU, 72.5 μL), and the soln was stirred for 2 h at 0 °C. The reaction mixture was directly chromatographed on a column of silica gel (40:1 CH_2Cl_2 –MeOH) to give **10** (0.5 g, quantitative) as an amorphous mass: $[\alpha]_{\text{D}} + 18.5^\circ$ (c 1.0, CHCl_3); ^1H NMR (CDCl_3): δ 1.70 (t, 1 H, $J_{\text{gem}} = J_{3\text{ax},4} = 10.83$ Hz, H-3dax), 1.98–2.31 (14 s, 42 H, 12 AcO and 2 AcN), 2.57 (dd, 1 H, $J_{3\text{eq},4} = 4.9$ Hz, H-3deq), 3.64 (dd, 1 H, $J_{2,3} = 9.9$, $J_{3,4} = 3.5$ Hz, H-3b), 3.87 (s, 3 H, MeO), 3.97 (d, 1 H, $J_{1,2} = 8.0$ Hz, H-1b), 4.95 (d, 1 H, $J_{1,2} = 9.1$ Hz, H-1c), 5.33 (dd, 1 H, $J_{6,7} = 3.6$, $J_{7,8} = 11.3$ Hz, H-7d), 5.42 (d, 1 H, $J_{3,4} = 3.5$ Hz, H-4b), 5.66 (m, 1 H, H-8d), 5.86 (dd, 1 H, $J_{2,3} = 10.8$, $J_{3,4} = 2.9$ Hz, H-3c), 6.45 (d, 1 H, $J_{1,2} = 3.6$ Hz, H-1a), 6.71 (d, 1 H, $J_{\text{NH},2} = 8.2$ Hz, NH), 8.63 (s, 1 H, C=NH). Anal. Calcd for $\text{C}_{58}\text{H}_{78}\text{Cl}_3\text{N}_3\text{O}_{36}$ (1499.6): C, 46.45; H, 5.24; N, 2.80. Found: C, 46.45; H, 5.15; N, 2.69.

2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-

galactopyranosyl-(2 \rightarrow 6)-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-2,4-di-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-acetyl- β -D-glucopyranosyl-(1 \rightarrow 1)-(2S,3R,4E)-2-azido-3-O-(tert-butyl-diphenylsilyl)-4-octadecene-1,3-diol (12).—To a soln of **10** (0.5 g, 0.33 mmol) and (2S,3R,4E)-2-azido-3-O-(tert-butyl-diphenylsilyl)-4-octadecene-1,3-diol (**11**; 0.37 g, 0.66 mmol) in dry CH_2Cl_2 (7.0 mL) was added molecular sieves (AW-300; 5 g), and the mixture was stirred for 5 h at room temperature, then cooled to 0 °C. To the cooled mixture was added trimethylsilyl trifluoromethanesulfonate (Me_3SiOTf ; 130 μL , 0.66 mmol), and the mixture was stirred for 12 h at 0 °C and then filtered. The insoluble materials were washed with CH_2Cl_2 , and the combined filtrate and washings was successively washed with M NaHCO_3 and water, dried (Na_2SO_4), and concd. Column chromatography (20:1 CH_2Cl_2 –MeOH) of the residue on silica gel gave **12** (50 mg, 80%) as an amorphous mass: $[\alpha]_{\text{D}} - 18.5^\circ$ (c 0.9, CHCl_3); ^1H NMR (CDCl_3): δ 0.88 (t, 3 H, $J_{\text{Me,CH}_2} = 6.6$ Hz, MeCH_2), 1.22 (s, 22 H, 11 CH_2), 1.66 (t, 1 H, $J_{\text{gem}} = J_{3\text{ax},4} = 12.6$ Hz, H-3dax), 1.88–2.29 (14 s, 42 H, 12 AcO and 2 AcN), 2.57 (dd, 1 H, $J_{3\text{eq},4} = 4.4$ Hz, H-3deq), 3.84 (s, 3 H, MeO), 3.96 (d, 1 H, $J_{1,2} = 6.4$ Hz, H-1b), 4.83 (d, 1 H, $J_{1,2} = 8.2$ Hz, H-1c), 4.95 (dd, 1 H, $J_{1,2} = 8.4$, $J_{2,3} = 9.5$ Hz, H-2a), 5.08 (t, 1 H, $J_{2,3} = J_{3,4} = 9.3$ Hz, H-3a), 5.17 (d, 1 H, $J_{\text{NH},2} = 5.3$ Hz, NH), 5.33 (dd, 1 H, $J_{6,7} = 3.6$, $J_{7,8} = 11.2$ Hz, H-7d), 5.42 (d, 1 H, $J_{3,4} = 3.5$ Hz, H-4b), 5.66 (m, 1 H, H-8d), 5.73 (dd, 1 H, $J_{2,3} = 8.6$, $J_{3,4} = 3.8$ Hz, H-3c), 6.58 (d, 1 H, $J_{\text{NH},2} = 7.7$ Hz, NH), 7.27–7.67 (m, 10 H, 2 Ph). Anal. Calcd for $\text{C}_{90}\text{H}_{129}\text{N}_5\text{O}_{37}\text{Si}$ (1901.1): C, 56.86; H, 6.84; N, 3.68. Found: C, 56.57; H, 6.83; N, 3.57.

2-Acetamido-3,4,6-tri-O-acetyl-2-deoxy- β -D-galactopyranosyl-(2 \rightarrow 6)-[(methyl 5-acetamido-4,7,8,9-tetra-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate)-(2 \rightarrow 3)]-2,4-di-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-acetyl- β -D-glucopyranosyl-(1 \rightarrow 1)-(2S,3R,4E)-3-O-(tert-butyl-diphenylsilyl)-2-octadecanamido-4-octadecene-1,3-diol (14).—To a soln of **12** (300 mg, 0.16 mmol) in benzene (13.5 mL) and water (0.6 mL) was added triphenylphosphine (84 mg, 0.32 mmol), and the mixture was stirred for 24 h at 30 °C and concd to give the crude amine **13**. A soln of the residue in dry CH_2Cl_2 (9 mL) was stirred with octadecanoic acid (136 mg, 0.48 mmol) and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (WSC; 92 mg, 0.48 mmol) for 24 h at room temperature.

Dichloromethane (120 mL) was added, and the soln was washed with water, dried (Na_2SO_4), and concd. Column chromatography (30:1 CH_2Cl_2 –MeOH) of the residue on silica gel gave **15** (236 mg, 70%) as an amorphous mass: $[\alpha]_{\text{D}} -14.7^\circ$ (c 0.9, CHCl_3); ^1H NMR (CDCl_3): δ 0.88 (t, 6 H, $J_{\text{Me,CH}_2}$ 6.4 Hz, 2 MeCH_2), 1.22 (s, 52 H, 26 CH_2), 1.85–2.30 (14 s, 42 H, 12 AcO and 2 AcN), 2.57 (dd, 1 H, $J_{3\text{eq},4}$ 4.8, J_{gem} 12.8 Hz, H-3deq), 4.97 (dd, 1 H, $J_{1,2}$ 7.8, $J_{2,3}$ 8.2 Hz, H-2a), 5.39 (d, 1 H, $J_{3,4}$ 3.3 Hz, H-4b), 5.66 (m, 1 H, H-8d), 5.73 (dd, 1 H, $J_{2,3}$ 11.5, $J_{3,4}$ 4.1 Hz, H-3c), 6.53 (d, 1 H, $J_{\text{NH},2}$ 6.7 Hz, NH), 7.27–7.73 (m, 10 H, 2 Ph). Anal. Calcd for $\text{C}_{108}\text{H}_{165}\text{N}_3\text{O}_{38}\text{Si}$ (2141.6): C, 60.57; H, 7.77; N, 1.96. Found: C, 60.42; H, 7.57; N, 1.88.

2-Acetamido-2-deoxy- β -D-galactopyranosyl-[(5-acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonic acid)-(2 \rightarrow 3)]-O-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 1)-(2S,3R,4E)-2-octadecanamido-4-octadecene-1,3-diol (**15**).—To a soln of **14** (300 mg, 0.16 mmol) in acetonitrile (5 mL) was added tetrabutylammonium fluoride (TBAF), and the soln was stirred for 24 h at 30 °C, and water (0.5 mL) was added. The soln was stirred for an additional 24 h at room temperature, then it was treated with Amberlite IR-120 (H^+) resin and filtered. The resin was washed with MeOH, and the combined filtrate and washings was concd. Column chromatography (5:5:1 CHCl_3 –MeOH– H_2O) of the residue on Sephadex LH-20 gave **15** (87 mg, quantitative) as an amorphous mass: $[\alpha]_{\text{D}} +30.0^\circ$ (c 1.0, 5:5:1 CHCl_3 –MeOH– H_2O); ^1H NMR [$(\text{CD}_3)_2\text{SO}$]: δ 0.88 (t, 6 H, $J_{\text{Me,CH}_2}$ 6.4 Hz, 2 MeCH_2), 1.22 (s, 52 H, 26 CH_2), 1.85 and 2.30 (2 s, 6 H, 2 AcN), 2.71 (dd, 1 H, J_{gem} 7.2, $J_{3\text{eq},4}$ 4.8 Hz, H-deq), 4.83 (d, 1 H, $J_{1,2}$ 6.0 Hz, H-1a), 4.89 (d, 1 H, $J_{1,2}$ 6.1 Hz, H-1c), 5.31, 5.51 (2 m, 2 H, H-4,5 for Cer unit). Anal. Calcd for $\text{C}_{66}\text{H}_{120}\text{N}_3\text{O}_{24}$ (1339.68): C, 59.17; H, 9.03; N, 3.14. Found: C, 58.90; H, 8.93; N, 3.12.

Acknowledgements

This work was supported in part by Grants-in-Aid for Scientific Research (B) (No. 07456162) and Scientific Research on Priority Areas (No. 05274102) from the Ministry of Education, Science and Culture of Japan, and a grant (NS 09626) from the United States National Institutes of Health.

References

- [1] Y. Makimura, H. Ishida, M. Kiso, and A. Hasegawa, *J. Carbohydr. Chem.*, 15 (1996) 1097–1119.
- [2] Y.-T. Li and S.-C. Li, in J.T. Dingle, R.T. Dean, and W. Sly (Eds), *Lysosomes in Biology and Pathology*, Elsevier, Amsterdam, 1984, pp. 99–117.
- [3] K. Sandhoff, K. Harzer, and W. Furst, in C.R. Scriber, A.L. Beaudet, W.S. Sly, and D. Valle (Eds), *The Metabolic and Molecular Basis of Inherited Disease*, Vol. 2, McGraw-Hill, New York, 1995, pp. 2427–2441.
- [4] Y.-T. Li, M.Y. Mazzotta, C.-C. Wan, R. Orth, and S.-C. Li, *J. Biochem.*, 248 (1973) 7512–7515.
- [5] K. Hotta, H. Ishida, M. Kiso, and A. Hasegawa, *J. Carbohydr. Chem.*, 13 (1994) 175–191.
- [6] M. Kiso and L. Anderson, *Carbohydr. Res.*, 72 (1979) c12–c14.
- [7] (a) T. Murase, H. Ishida, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, 184 (1988) C1–C4; (b) A. Hasegawa, T. Nagahama, H. Ohki, K. Hotta, H. Ishida, and M. Kiso, *J. Carbohydr. Chem.*, 10 (1991) 493–498.
- [8] A. Marra and R. Sinaÿ, *Carbohydr. Res.*, 187 (1989) 35–42.
- [9] (a) G.H. Veeneman, S.H. van Leeuwen, and J.H. van Boom, *Tetrahedron Lett.*, 31 (1990) 1331–1334; (b) P. Konradsson, D.R. Mootoo, R.E. McDevitt, and B. Fraser-Reid, *J. Chem. Soc., Chem. Commun.*, (1990) 270–272.
- [10] J.F.G. Vliegthart, L. Dorland, H. Van Halbeek, and J. Haverkams, in R. Schauer (Ed.), *Sialic Acids; Chemistry, Metabolism, and Function, Cell Biology Monographs*, Vol. 10, Springer, Vienna, New York, 1982, pp. 127–172.
- [11] K. Jansson, S. Ahlfors, T. Frejd, J. Kihlberg, G. Magnusson, J. Dahmen, G. Noori, and K. Stenvall, *J. Org. Chem.*, 53 (1988) 5629–5647.
- [12] (a) M. Numata, M. Sugimoto, K. Koike, and T. Ogawa, *Carbohydr. Res.*, 163 (1987) 209–225; (b) R.R. Schmidt and J. Michel, *Angew. Chem., Int. Ed. Engl.*, 19 (1980) 731–732.
- [13] A. Hasegawa, T. Nagahama, H. Ohki, and M. Kiso, *J. Carbohydr. Chem.*, 11 (1992) 699–714.
- [14] T. Ehara, A. Kameyama, Y. Yamada, H. Ishida, M. Kiso, and A. Hasegawa, *Carbohydr. Res.*, 281 (1996) 237–252.
- [15] M. Mori, Y. Ito, and T. Ogawa, *Carbohydr. Res.*, 195 (1990) 199–224.
- [16] K.C. Nicolaou, T. Caulfield, H. Kataoka, and T. Kumazawa, *J. Am. Chem. Soc.*, 110 (1988) 7910–7912.
- [17] T. Fujisawa, T. Mori, K. Fukumoto, and T. Sato, *Chem. Lett.*, (1982) 1891–1894.